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Transmitted herewith for filing under 37 CFR 1.53(b) is the

- ☒ patent application of  
☐ continuation patent application of  
☐ divisional patent application of  
☐ continuation-in-part patent application of

Inventor(s)/Applicant Identifier: Ping Kuen Lam and Eric Sun-Yin Chan

For: MATERIAL AND METHOD FOR ENGRAFTMENT OF A COMPOSITE BIOCOMPATIBLE SKIN GRAFT ON THE NEODERMIS OF ARTIFICIAL SKIN

☐ This application claims priority from each of the following Application Nos./filing dates:

the disclosure(s) of which is (are) incorporated by reference.

☐ Please amend this application by adding the following before the first sentence: "This application is a ☐ continuation ☐ continuation-in-part of and claims the benefit of U.S. Application No. 60/\_\_\_\_\_, filed \_\_\_\_\_, the disclosure of which is incorporated by reference."

Enclosed are:

☒ 14 pages of specification

☒ 3 pages of claims

☒ 1 page of Abstract

☒ 4 pages of Appendix

☒ 2 sheets of formal drawings.

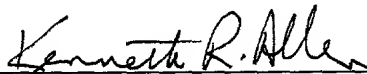
☒ 1 page Recordation Form Cover Sheet.

☒ An assignment of the invention to The Chinese University of Hong Kong

☒ A signed Declaration & Power of Attorney

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**PATENT APPLICATION**  
**MATERIAL AND METHOD FOR ENGRAFTMENT OF A COMPOSITE**  
**BIOCOMPATIBLE SKIN GRAFT ON THE NEODERMIS OF**  
**ARTIFICIAL SKIN**

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**MATERIAL AND METHOD FOR ENGRAFTMENT OF A  
COMPOSITE BIOCOMPATIBLE SKIN GRAFT ON THE  
NEODERMIS OF ARTIFICIAL SKIN**

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REFERENCE TO SPONSORSHIP

This invention was made under the joint sponsorship of the Chinese University of Hong Kong of Shatin, Hong Kong SAR, and the Hospital Authority, also known as the Prince of Wales Hospital of Shatin, Hong Kong SAR.

BACKGROUND OF THE INVENTION

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This invention relates to the cultivation of keratinocytes and dermal fibroblasts on a biosynthetic membrane and subsequent engraftment of this type of membrane on the neodermis of artificial skin with particular application to humans. This invention also relates to fabricated graft materials.

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Prompt wound coverage for protection and stabilization is essential for the treatment of burns. The 3T3 cell-feeder layer technique developed by Rheinwald and Green in the 1970s is the standard method for cultivation of autologous epidermal autograft or so-called cultivated epidermal autograft material ("CEA"). A small piece of native skin from the patient can be cultured and expanded to 500 times or more in size within 3-4 weeks. However, the lack of dermis and the fragility of the cultured graft often result in unpredictable grafting success rates ranging from 0%-80%, as reported by Tompkins et al. (1992) and Herzog et al. (1991).

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Alloderm<sup>TM</sup> and Integra<sup>TM</sup> are two currently popular examples of human skin substitute commercially available in the market. Integra<sup>TM</sup>, a brand of artificial skin is sold by Integra LifeScience Corporation of New Jersey, USA, and has been approved by FDA for use in the USA since 1996. Artificial skin is a bilayer biosynthetic sheet comprising porous collagen-glycoaminoglycan integrated with a thin silicone membrane as an outer layer. The use of artificial skin such as Integra as a biocompatible acellular dermal replacement in deep and full-thickness burn wounds is well known.

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It has been observed that within about 14 to 21 days following the grafting of Integra, there is full vascularization of the neodermis formed in the Integra. Thereafter an ultra thin split thickness skin graft must be harvested from a donor site in order to cover the neodermis immediately after the silicone membrane is removed. Substantial research effort has been undertaken in the past to determine the possibility of reliably

grafting CEA on the neodermis, since an effective combination of CEA and Integra should eliminate the second operative stage, the associated pain and scarring, as well as a need for a second donor site, which may not be available in extensively burned patients.

If the grafted CEA does not 'take' on the neodermis of Integra after the silicone

5 membrane is peeled off, it can be replaced by another CEA. Whereas, in the conventional application of Integra, another split thickness autograft must be harvested from a second or even a third donor site. There have been very limited initial anecdotal reports on experience with such a combination technique, such as Sheridan et al. 1999 and Pandya et al. 1998. At the 10th Congress of International Society For Burn Injuries, November  
10 1998 in Israel, the difficulties with the conventionally cultured graft anchoring onto the neodermis of Integra were addressed. The exact reasons for such difficulties remain unknown.

Laserskin<sup>TM</sup> material is a thin and pliable biosynthetic membrane comprising a 100% benzyl esterified hyaluronic acid derivative suitable for use as a  
15 substratum in the growth of skin cells. The recommendation of the manufacturer is to seed human keratinocytes on Laserskin preseeded with irradiated 3T3 cells as feeder layer. When following the manufacturer's recommendation, it was found that, after the initiation of the formation of keratinocyte colonies, the xenogenic 3T3 cells growing on the Laserskin were less likely to be washed away than those growing on a culture dish as  
20 in the conventional Green's method during each flushing procedure with phosphate-buffered saline. It is believed that the remaining 3T3 cells or debris might have sensitized the host to xenogenic antigen resulting in undesired late graft rejection. What is needed is a cultivation and engraftment procedure with a biocompatible, durable human skin substitute.

#### 25 RELATED REFERENCES

The following references, not all of which are prior art for the purposes of a patent application, are hereby made of record and incorporated herein by reference for the purposes described in this text:

1) Rheinwald J, Green H. "Serial cultivation of strain of human epidermal  
30 keratinocytes: The formation of keratinizing colonies from single cells." **Cell** 1975; 6: 331-344.

2) Tompkins RG, Burke JF. "Burn wound closure using permanent skin replacement material." **World J Surgery** 1992; 16: 47-52.

3) Herzog SR, Meyer A, Woodley D, Peterson HD. "Wound coverage with cultured autologous keratinocytes: Use after burn wound excision, including biopsy follow-up." **J Trauma** 1991; 28: 195-199.

4) Yannas IV, Burke JF, Orgill DP, Skrabut EM. "Wound tissues can utilize a polymeric template to synthesize a functional extension of skin." **Science** 1982; 215: 174-176.

5) Heimbach D, Letterman A, Burke J et al. "Artificial dermis for major burns. A multi-center randomized clinical trial." **Ann Surgery** 1988; 208: 313-320.

6) Sheridan RL, Heggerty M, Tompkins RG, Burke JF. "Artificial skin in massive burns-results at ten years." **Eur J Plast Surgery** 1994; 17: 91-93.

7) Sheridan RL, Tompkins RG. "Skin substitutes in burns." **Burns** 1999; 25: 97-103.

8) Pandya AN, Woodward B, Parkhouse N. "The use of cultured autologous keratinocytes with Integra in the resurfacing of acute burns." **Plast Reconst Surgery** 1998; 102: 825-828.

9) Hultman CS, Brinson GM, Silitharm S, et al. "Allogenic fibroblasts used to grow cultured epidermal autografts persist in vivo and sensitize the graft recipient for accelerated second-set rejection." **J Trauma** 1996; 41: 51-60.

10) Lam PK, King WK, et al. "Development and evaluation of a new composite Laserskin graft" **Abstracts of the 10th Congress of International Society For Burn Injuries**, November 1-6 1998, Jerusalem, Israel, p. 46.

11) Sato T, Kirimura Y, Mori Y. "The co-culture of dermal fibroblasts with human epidermal keratinocytes induces increased prostaglandin E2 production and cyclooxygenase 2 activity in fibroblasts." **J Investigative Dermatology** 1997; 109: 334-339.

12) Pano RJ, Rubin JR, Aaronson SA, Mason R. "Keratinocytes growth factor and hepatocyte growth factor/scatter factor are heparin-binding growth factors for alveolar type II cells in fibroblast-conditioned medium." **J Clin Invest** 1993; 92: 969-977.

13) Burd DAR, Greco RM, Regauer MT, et al. "Hyaluron and wound healing: a new perspective." **Br J Plast Surgery** 1991; 44: 579-584.

14) Feinberg RN, Beebe DC. "Hyaluronate in vasculogenesis." **Science** 1983; 220: 1177-1179.

15) Alman DG. **Practical Statistics for Medical Research** Chapman & Hall, London 1991 pp. 211, 260-261.

16) Harris PA, Francesco F di, Barisoni D, Leigh IM, Navsaria HA. "Use of hyaluronic acid and cultured autologous keratinocytes and fibroblasts in extensive burns." **Lancet** 1999; 353: 35-36.

17) Myers SR, Grady J, Soranzo C, Sander R, et al. "A hyaluronic acid membrane delivery system for cultured keratinocytes: Clinical take rates in the porcine Kerato-Dermal model." **J Burn Care Rehabil** 1977; 18: 214-222.

18) Hansbrough JF, Dore C, Hansbrough WN. "Clinical trials of living dermal tissue replacement placed beneath meshed, split-thickness skin grafts on excised burn wounds." **J Burn Care Rehabil** 1992; 13: 519-529.

#### SUMMARY OF THE INVENTION

According to the invention, autologous cultured keratinocytes grown on a biocompatible substratum are engrafted on the neodermis of artificial skin covering a wound. Autologous keratinocytes may be cultivated on a commercially available membrane such as Laserskin<sup>TM</sup> (available from Fidia Advanced Biopolymers Ltd., Abano Terme (PD), Italy) following pre-seeding with autologous or allogenic dermal fibroblasts. The resultant composite material may then be applied on the neodermis of artificial skin which had been previously engrafted on the patient. The composite material, and specifically Composite Biocompatible Skin Graft (CBSG) material comprises autologous keratinocytes and allogenic or autologous dermal fibroblasts grown on the substratum. A method for fabricating the composite material includes the application of dermal fibroblasts onto the substratum as a feeder layer and then inoculating autologous keratinocytes on the resultant structure. A method for engraftment comprises first applying an artificial skin with a protective silicone membrane on a wound area, thereby allowing vascularization; following vascularization, removing the silicone membrane and engrafting the cultured composite material onto the vascularized artificial skin.

Human fibroblasts used in the cultivation technique according to the invention were found to achieve a role similar to 3T3 cells in the initiation of keratinocyte colonies on Laserskin. Specifically, it was found that the seeding efficacy of human keratinocytes was increased to up to 95%. CBSG containing autologous keratinocytes and autologous dermal fibroblast or allogenic dermal fibroblasts or a combination of autologous and allogenic dermal fibroblasts according to the invention, has been

successfully applied to burn patients whose wounds were previously grafted with allografts employing the engraftment techniques according to the invention.

5 The purpose of this invention is to simplify burn treatment further and eventually to save lives of patients having extensive burns where little or no autologous skin grafts can be repeatedly harvested in a short period of time. With the inventive technique, all dead skin tissue of a patient with extensive burns can be excised within about three to seven days after injury. The wound can be covered with artificial skin, such as Integra<sup>TM</sup> or any dermal equivalent thereof, and only a small quantity of harvested normal skin would be required to initiate skin culture, which may thereafter be engrafted, according to the invention, on the neodermis of the artificial skin, with resultant lower rejection and infection incidences.

CBSG material according to the invention offers notable advantages. First, the basal proteins (including the early basement membrane proteins such as collagen IV and fibronectin) of the cultured graft are protected from dispase treatment because the keratinocytes are directly cultivated on the pliable Laserskin. This is believed to enhance anchorage of the cultured keratinocytes on the neodermis of Integra. Second, in addition to acting as a feeder layer, the dermal fibroblasts in the inventive CBSG material evidently produce a number of proteins such as native collagen fibers and fibronectin which is believed to facilitate the attachment of a cultured graft. Third, the cultured keratinocytes of the inventive CBSG can be grafted five to seven days sooner than can traditionally-cultured keratinocytes. This is because cultured keratinocytes of the inventive CBSG are capable of being transferred and grafted at the sub-confluent or less differentiated stage rather than at a later confluent stage. Fourth, since there is minimal need for a donor site there is less likelihood of widespread scarring related to donor site harvesting. Fifth, the cultured keratinocytes of CBSG can be handled much more easily than the conventional CEA during its application on the neodermis of the artificial skin. Fewer cultured cells are lost or damaged during the transfer and application of CBSG. This should improve the success rate of the cultured graft. Sixth, the inventive engraftment technique can result in higher demand and broader scope of clinical applications for artificial skin.

The invention will be better understood by reference to the following detailed description in connection with the accompanying drawings.

## BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1A-1C are schematic diagrams of cross-sections of Type A, B and C embodiments of a cultured graft material according to the invention.

Fig. 2A-2C are schematic diagrams of cross-sections of skin grafts illustrating the engraftment technique according to the invention.

## DESCRIPTION OF THE SPECIFIC EMBODIMENTS

- Fig. 1A, is what is herein termed a Type A composite skin graft material. In a specific embodiment it is formed by cultivating autologous keratinocytes on a commercially-available biosynthetic substratum 14 of a derivative of benzyl esterified hyaluronic acid. On the basal side and on the upper side of the substratum 14 layers of dermal fibroblasts 12 are seeded and grown. Over the dermal fibroblasts 12 of the upper side, a layer of autologous keratinocytes 10 is grown. The keratinocytes 10 are harvested from the target donor patient. The dermal fibroblasts can be autologous or allogenic.
- Fig. 1B is a Type B composite skin graft material. In a specific embodiment it is formed of a cultivation of autologous keratinocytes on commercially-available biosynthetic substratum 14. On the upper side of the substratum 14 there is a layer of dermal fibroblasts 12. Over the dermal fibroblasts 12 of the upper side is a layer of autologous keratinocytes 10. The dermal fibroblasts can be autologous or allogenic.
- Fig. 1C is a Type C composite skin graft material. Originally intended as a control, it is, in a specific embodiment, formed of a cultivation of autologous keratinocytes on substratum 14. On the upper side of the substratum 14 there is merely a layer of keratinocytes 10.

- Fig. 2A is an illustration of a first stage in an engraftment method according to the invention. In Fig. 2A there is a wound bed 26 of a patient on Day Zero prior to the commencement of treatment. An artificial skin substrate 20 is applied upon the wound bed 26. The artificial skin substrate 20 has a basal side and an upper side. It comprises a layer 24 of collagen-glycoaminoglycan on its basal side, thus in a position to be juxtaposed to the wound bed 26. A covering membrane 22 of silicone is on the opposing upper side of the substrate 20. For a period of about fourteen days to about



twenty-one days, the wound bed remains covered in order to allow vascularization of the neodermis. Referring to Fig. 2B, at this point, the silicone membrane 22 is removed from the vascularized neodermis of artificial skin 24' in preparation for immediate further treatment. Referring to Fig. 2C, one of Type A, B or C composite skin graft material 28 is engrafted on the layer 24'. In the specific embodiment, the material 28 comprises a substratum 34 having thereon autologous cultivated keratinocytes 30 of the donor patient, typically having thereunder on the substratum a layer of dermal fibroblasts 32, depending on the type of graft material 28.

The following is a detailed discussion of the materials and results of actual clinical experiments.

### **Laserskin**

Laserskin is a biosynthetic biocompatible substratum for keratinocyte cultivation according to the invention. It is a form of thin and pliable biosynthetic membrane comprising a 100% benzyl esterified hyaluronic acid derivative. Laserskin is commercially available from Fidia Advanced Biopolymers Ltd., Abano Terme (PD), Italy. Laserskin was found to be useful to the inventors' experiments, although its preparation and application are not done according to the manufacturer's conventional instructions. There is nothing to preclude the use of compatible bioequivalents or human skin substitutes would also work in a similar fashion with the inventive CBSG.

### **Keratinocyte and fibroblast culture sources**

In the specific experiments undertaken, human keratinocytes and fibroblasts were derived from human foreskin obtained from circumcision for the human part of study. Rat keratinocytes and fibroblasts were obtained from Sprague Dawley rat (body weight, 400-450 gm). The primary cultures of human and rat keratinocytes were grown on mitomycin C-treated 3T3 feeder layer. The secondary and tertiary cultures of human keratinocytes were maintained in defined keratinocyte growth medium (Gibco), whereas rat keratinocytes were continuously grown on 3T3 feeder layer. Fibroblasts were cultivated in DMEM supplemented with 10% FBS.

### **Comparison of seeding efficacy of human keratinocytes on Laserskin**

Human dermal fibroblasts and 3T3 cells were grown separately on Laserskin with DMEM supplemented with 10% FBS. At sub-confluency, the 3T3 cells were treated with mitomycin-C (4mg/ml) for 2 hours at 37 °C, whereas the human fibroblasts were not treated. Human keratinocytes were seeded ( $3 \times 10^4$  cells/cm<sup>2</sup>) on top of the Laserskin either pre-seeded with 3T3 cells or human fibroblasts. The keratinocyte suspension was instilled and concentrated on the Laserskin surface (0.5ml/cm<sup>2</sup>). Thirty minutes after seeding, DMEM supplemented with 10% FBS, insulin (4mg/ml), cholera toxin (6ng/ml), and EGF (10ng/ml) (per Rheinwald et al. 75) were added to the culture.

In a control run, human keratinocytes were also seeded on top of plain Laserskin (Figure 1C). Efficacy was exhibited as hereinafter noted. After incubation at 37 °C in a humidified atmosphere with 5% CO<sub>2</sub> for 24 hours, the non-seeded keratinocytes on the Laserskin were gently rinsed off with PBS. The cells were then counted with a hemocytometer. In order to quantify the number of fibroblasts or 3T3 cells detached together with the non-seeded keratinocytes during the washing of the Laserskin, the Laserskin without keratinocytes but seeded with equivalent number of 3T3 cells or fibroblasts were included as control.

#### **Comparison of seeding efficacy of rat keratinocytes on Laserskin**

The seeding of rat keratinocytes on Laserskin was performed according to a technique essentially the same as the seeding of human keratinocytes. Rat fibroblasts were used to replace the human fibroblasts.

#### **Preparation of CBSG graft for use on full-thickness wounds**

Three different types of composite skin graft (CBSG) materials were compared and tested in animals, specifically laboratory rats. Type A CBSG consisted of allogenic rat fibroblasts seeded onto the basal side of a Laserskin substratum. The fibroblasts were stimulated to produce collagen and other proteins by feeding the fibroblasts with DMEM supplemented with 10% FBS and 50µm ascorbic acid. After ten days, the Laserskin substratum was turned over and the upper side was seeded with fibroblasts. The fibroblasts on both sides were then fed with DMEM supplemented with 10% FBS. When the fibroblasts on the upper side became sub-confluent, rat keratinocytes were seeded ( $3 \times 10^4$  cells/cm<sup>2</sup>) on top of the upper surface of CBSG material. The cells were fed with DMEM supplemented with 10% FBS, insulin

(4mg/ml), cholera toxin (6ng/ml), EGF(10ng/ml). The keratinocytes became sub-confluent and were ready for grafting after only four to six days.

Two further types of CBSG materials were prepared. In Type B CBSG materials, the fibroblasts and keratinocytes were grown only on the upper side of the substratum. In Type C CBSG, keratinocytes alone were seeded on the upper side. As will be evident hereafter, all three types had a demonstrable efficacy.

### **Grafting Procedures:**

Using twenty Sprague Dawley rats for the experiment, three separate full-thickness wounds, 2 cm in diameter each, were created on the dorsal area of each animal. The CBSG sheets were rinsed thoroughly with sterile PBS in a culture dish. They were lifted from the culture dish and placed directly on the wound so that the basal side of the CBSG was in contact with the wound. Non-adherent dressing was placed over the graft site. Each animal was engrafted with the three different types of CBSG materials. The migration of the epithelium from the cut wound edge was prevented by application of a polypropylene ring. The entire dressing was protected with porous thermal plastic. All grafts consisted of autologous keratinocytes and allogenic dermal fibroblasts.

### **Data Analysis:**

The 'take' rate of the CBSG materials was analyzed with the Fisher Exact Test using Stat Xact (version 2.05). In multiple comparisons, the p values were further adjusted by the Bonferron method (i.e., multiplying the p value obtained from each test by k paired comparisons) in order to account for the so called 'serial t test'.

### **Results**

Twenty-four hours after the seeding on the CBSG substratum, the non-viable and unseeded keratinocytes were carefully washed away and concentrated for counting. There were no detectable 3T3 cells or fibroblasts detached from the Laserskin during the washing procedure. The seeding efficacy of keratinocytes was calculated as:

$$\frac{[\text{Total keratinocytes count (before seeding)} - \text{non-seeded keratinocytes}]}{\text{Total keratinocytes count (before seeding)}} \times 100\%$$

It is evident that the selected type of CBSG substratum provided a suitable culture template for the in vitro proliferation of keratinocytes. On a plain Laserskin substratum, human keratinocytes showed a mean seeding efficacy of 75%. These human keratinocytes had a 95% seeding efficacy on Laserskin populated with human fibroblasts and a 98% on 3T3 cell-seeded Laserskin (Table 1). Rat keratinocytes had a seeding efficacy of 36% on plain Laserskin. The respective seeding efficacies of rat keratinocytes on 3T3/Laserskin and on allogenic fibroblasts/Laserskin were 91% and 88%. The seeding efficacies of human/rat keratinocytes growing on 3T3 cell/Laserskin or on allogenic fibroblasts/Laserskin were significantly ( $p < 0.001$ ) better than those seeded on plain Laserskin, as noted in the Fisher Exact Testing using Stat Xact (version 2.05) statistical package.

It is believed that human/rat fibroblasts could achieve a role similar to that of the 3T3 cells in enhancing the seeding efficacies of keratinocytes growing on Laserskin with respective  $p$  values of 0.445 and 0.646 using the same statistical package.

The optical transparency of the Laserskin allowed regular inspection of the grafted wound bed as it healed. Skin biopsies were taken from the center of the grafted area of the subject. It was observed that the polypropylene ring prevented the migration of epithelium from the wound edge as no epithelial cell was found in the control rat wound sutured with polypropylene ring alone up to day 21. In sixteen out of the twenty (80%) animal wounds covered with Type A CBSG, the keratinocytes formed a multi-layered epithelium that had a basal layer in contact with underlying connective tissue. The undersurface of epidermis did not show rete. Fibrovascular ingrowth of connective tissue into the Laserskin was observed. Eight (40%) of the Type B CBSG sites and seven (35%) of the Type C CBSG sites showed re-epithelialization. Histologically there was no observable epithelium in 3 (15%), 10 (50%), and 12 (55%) of the grafted CBSG sites of Types A, B and C respectively (Table 2).

The graft take rate of Type A CBSG was found to be significantly higher than those of Types B and C CBSG in the Fisher Exact Test/Bonferroni method analysis with respective  $p$  values of 0.0419 and 0.018. There was no significant difference ( $p > 0.1$ ) of the graft take rate between Types B and C CBSG (table 2). Infection occurred in only one (5%) animal with a Type A or C CBSG graft and in two (10%) animals with a Type B CBSG graft.

A preliminary clinical trial of CBSG was also conducted on two human burn patients (designated CLB & CKT). Three out of the five (60%) grafted wounds in patient CLB developed stratified epithelium (Table 3). The other two grafts were lost because of the bacterial colonization at the adjacent tissues. All three skin biopsies from patient CKT showed complete epithelization with primitive rete ridge formation. There was no severe immunologic reaction to the allogenic fibroblasts in these two patients or in the animal experiments. Clinically, the CBSG grafts were durable. All grafts were histologically confirmed as a 'take' at two weeks, and they remained clinically intact on the recipient wound at eight weeks post-grafting. No further biopsies were taken of the patients.

## Discussion

Problems encountered in cultivation of keratinocytes on Laserskin type material can be attributed to limited experience on this new product. Hyaluronic acid is a mucopolysaccharide with alternating  $\beta$  1-3 glucuronidic and  $\beta$  1-4 glycosaminidic bonds. Laserskin mainly consists of 100% benzyl esterified hyaluronic acid. A weak electrostatic association exists between the keratinocytes and the Laserskin. Since the volume and surface area of human keratinocytes differ from those of rat keratinocytes, there is a variation of charge density of two cell populations. Consequently they have different seeding efficacies on plain Laserskin. The species-specific difference was abolished by cultivating keratinocytes on 3T3 cell/Laserskin because 3T3 cells were the dominant factors influencing keratinocyte attachment on Laserskin.

The seeding efficacies of human and rat keratinocytes ( $3 \times 10^4$  cells/cm<sup>2</sup>) were 75% and 36% respectively on plain Laserskin. The manufacturer recommends seeding the human keratinocytes on Laserskin pre-seeded with irradiated 3T3 cells. Good seeding efficiencies of 98% (human keratinocytes) and 91% (rat keratinocytes) were demonstrated on Laserskin with Mitomycin-treated 3T3 cells. In Green's method, keratinocytes are seeded on a 3T3 cell feeder layer which allows a rather low seeding density of  $6-10 \times 10^4$  cells/cm<sup>2</sup> for the primary culture. A smaller seeding density of  $2-5 \times 10^4$  cells/cm<sup>2</sup> is feasible for the secondary and tertiary cultures. The utility of cultured epidermal autograft grown on culture dish using Green's feeder layer technique is limited by the persistent 3T3 fibroblasts which sensitize the host to xenogenic antigen resulting in late graft rejection. The 3T3 cells were found less likely to be washed away from the

Laserskin. However, allogenic fibroblasts can achieve a similar role as 3T3 cells in initiation of keratinocyte colonization on Laserskin.

It is evident that the CBSG material consisting of cultured keratinocytes and dermal fibroblasts is a good human skin substitute for freshly excised full-thickness wounds which were previously grafted with allografts.

	Human Keratinocytes	Rat keratinocytes
Plain Laserskin	75%	36%
Laserskin with 3T3 cells	98%	91%
Laserskin with human fibroblasts	95%	-
Laserskin with rat fibroblasts	-	88%

**Table 1** Mean Seeding Efficacies of Human and Rat Keratinocytes on Laserskin. 15 determinations were made. Seeding efficacies of human/rat keratinocytes were significantly improved by 3T3 cells/fibroblasts ( $p < 0.001$ ).

Results	Type A(N=20)	Type B(N=20)	Type C(N=20)
	Keratinocytes + Fibroblasts on both sides	Keratinocytes + Fibroblasts on upper side	Keratinocytes alone
Grafted epithelium observed	16 (80%)	8 (40%)	7 (35%)
No epithelium observed	3 (15%)	10 (50%)	12 (55%)
Infection	1(5%)	2 (10%)	1 (5%)

**Table 2** Evaluation of Composite Biocompatible Skin Graft on Animals  
Type A CBSG had a higher graft take rate than types B & C with respective  $p$  values of 0.0419 and 0.018.

	Grafted sites	Graft Histology
	Left hand (2)	v v
Patient CLB	Left arm	X
	Left forearm	v
	Left shoulder	X
Patient CKT	Left arm	v
	Left forearm (2)	v v

**Table 3** Preliminary Clinical Trials on Composite Biocompatible Skin Graft.  
'v' = 'take'; 'X' = 'not take'

Attempts were made to graft freshly harvested epidermis onto the neodermis of Integra in animal experiments. The dermal-epidermal separation was performed with dispase. The epidermal graft was not easily 'taken' by the neodermis whereas thin split thickness autografts readily incorporated on the neodermis as in many clinical applications. It is believed that the presence of fibroblasts in the dermis of the thin autograft may play an essential role in the keratinocyte attachment and its proliferation on the neodermis. It was observed that, during wound healing, cell-cell interactions between epidermal keratinocytes and dermal fibroblasts contributed to the organization of epidermis. Prostaglandin E2 (PGF2) is considered to be involved in the proliferation and differentiation of keratinocytes. The production of PEG2 was enhanced in the co-culture of keratinocytes and fibroblasts, whereas the PEG2 production was negligible in monolayer cultures of keratinocytes or fibroblasts. The fibroblasts also produced two soluble heparin-binding growth factors such as keratinocyte growth factor and hepatocyte growth factor/scatter factor that promote DNA synthesis and proliferation of keratinocytes.

The combination of the inventive CBSG material including keratinocytes and dermal fibroblasts and Integra artificial skin was applied to a 28-year old man with extensive hypertrophic burn scars and neck contracture. Multiple wounds from scar

excision with a total area of approximately 250 cm<sup>2</sup> were covered with Integra. At the same time a skin biopsy was taken for keratinocyte culture. Two weeks later, the top silicone layer of the artificial skin was removed. The neodermis of the artificial skin was then covered with the CBSG. At Days 6 and 14 after the application of CBSG, biopsies  
5 were taken from center of one of the wounds. The histology at Day 6 showed early epithelization with a few squamous epithelial cells settling on top of the scaffolds of Integra and Laserskin. Most of the epithelium were single-layer flattened squamous cells. The biopsy taken on Day 14 showed a viable 2- to 4-cell-thick squamous epithelium with minimal inflammation. The wounds were completely epithelized within four weeks of  
10 the application of the CBSG. Biopsies taken from other wounds showed similar findings.

The invention has now been explained with reference to specific embodiments. Other embodiments will be evident to those of ordinary skill in the art. It is therefor not intended that this invention be limited, except as indicated by the appended claims.



WHAT IS CLAIMED IS:

1                   1.       A method for cultivation of graftable skin comprising:  
2                   growing a layer of dermal fibroblasts upon at least an upper side of a  
3       biosynthetic substratum of a derivative of benzyl esterified hyaluronic acid; and, after  
4       said dermal fibroblast layer becomes at least sub-confluent,  
5                   growing a layer of keratinocytes over said dermal fibroblasts upon said  
6       upper side of said substratum to form a composite skin graft material, said keratinocytes  
7       being harvested from a target donor patient.

1                   2.       The method according to claim 1 wherein said dermal fibroblasts  
2       are allogenic.

1                   3.       The method according to claim 1 wherein said dermal fibroblasts  
2       are autologous.

1                   4.       A method for cultivation of graftable skin comprising:  
2                   growing a first layer of dermal fibroblasts upon a first basal side of a  
3       biosynthetic substratum of a derivative of benzyl esterified hyaluronic acid;  
4                   growing second layer of dermal fibroblasts upon a second upper side of  
5       said biosynthetic substratum; and  
6                   after said second dermal fibroblast layer becomes at least sub-confluent,  
7       growing a layer of keratinocytes over said dermal fibroblasts upon said upper side of said  
8       substratum to form a composite skin material, said keratinocytes having been harvested  
9       from a target donor patient.

1                   5.       The method according to claim 4 wherein said dermal fibroblasts  
2       are allogenic.

1                   6.       The method according to claim 4 wherein said dermal fibroblasts  
2       are autologous.

1                   7.       A method for cultivation of graftable skin comprising:  
2                   growing a layer of keratinocytes upon an upper side of a substratum of a  
3       biosynthetic substratum of a derivative of benzyl esterified hyaluronic acid to form a

4 composite skin graft material, said keratinocytes having been harvested from a target  
5 donor patient.

1 8. A graftable skin material comprising a composite of:  
2 a biosynthetic substratum of a derivative of benzyl esterified hyaluronic  
3 acid;  
4 a layer of dermal fibroblasts upon at least an upper side of said  
5 biosynthetic substratum; and  
6 a layer of keratinocytes over said dermal fibroblasts upon said upper side  
7 of said substratum, said keratinocytes having been harvested from a target donor patient.

1 9. The material according to claim 8 wherein said dermal fibroblasts  
2 are allogenic.

1 10. The material according to claim 8 wherein said dermal fibroblasts  
2 are autologous.

1 11. A graftable skin material comprising a composite of:  
2 a biosynthetic substratum of a derivative of benzyl esterified hyaluronic  
3 acid;  
4 a first layer of dermal fibroblasts upon a first basal side of said  
5 biosynthetic substratum;  
6 a second layer of dermal fibroblasts upon a second upper side of said  
7 biosynthetic substratum; and  
8 a layer of keratinocytes over said dermal fibroblasts upon said upper side  
9 of said substratum, said keratinocytes having been harvested from a target donor patient.

1 12. The material according to claim 11 wherein said dermal fibroblasts  
2 are allogenic.

1 13. The material according to claim 11 wherein said dermal fibroblasts  
2 are autologous.

1 14. A graftable skin material comprising:  
2 a substratum of a biosynthetic substratum of a derivative of benzyl  
3 esterified hyaluronic acid; and

4 a layer of keratinocytes upon an upper side of said substratum, said  
5 keratinocytes having been harvested from a target donor patient.

1 15. A method for grafting a graftable skin material comprising the  
2 steps of:

3 applying an artificial skin substrate upon a wound bed of a recipient  
4 patient; said artificial skin substrate comprising a layer of collagen-glycoaminoglycan on  
5 a basal side to be juxtaposed to said wound bed and a covering membrane of silicone on  
6 an opposing upper side;

7 allowing a sufficient time to form a vascularized wound bed under said  
8 collagen-glycoaminoglycan; thereupon

9 removing said silicone membrane; and thereupon

10 applying a basal side of a sheet of cultivated skin material over said  
11 collagen- glycoaminoglycan, said cultivated skin material comprising at least a layer of  
12 keratinocytes upon an upper side of a substratum, said keratinocytes being harvested from  
13 a target donor patient.

1 16. The method according to claim 15 wherein said cultivated skin  
2 material further comprises a layer of dermal fibroblasts upon at least an upper side of a  
3 biosynthetic substratum and wherein said layer of keratinocytes is over said dermal  
4 fibroblasts.

1 17. The method according to claim 16 wherein said cultivated skin  
2 material further comprises a layer of dermal fibroblasts upon said basal side of said  
3 biosynthetic substratum.

**MATERIAL AND METHOD FOR ENGRAFTMENT OF A COMPOSITE  
BIOCOMPATIBLE SKIN GRAFT ON THE NEODERMIS OF ARTIFICIAL SKIN**

**ABSTRACT OF THE DISCLOSURE**

Autologous cultured keratinocytes are grown on a biosynthetic and

- 5 biocompatible substratum following pre-seeding with autologous or allogenic dermal fibroblasts. The resultant composite material may then be applied on the neodermis of artificial skin which had been previously grafted on the patient. The composite material, and specifically Composite Biocompatible Skin Graft (CBSG) material comprises autologous keratinocytes and allogenic or autologous dermal fibroblasts grown on Laserskin. A method
- 10 for cultivating the CBSG includes the application of dermal fibroblasts onto the substratum as a feeder layer and then the inoculation of autologous keratinocytes on the resultant structure. A method for engraftment comprises first applying an artificial skin with a protective silicone membrane on a wound area, thereby allowing vascularization; and following vascularization, removing the silicone membrane and engrafting the CBSG material onto the
- 15 vascularized artificial skin.

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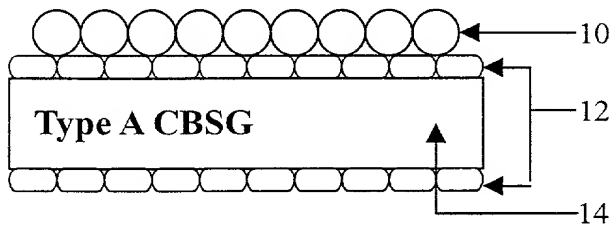


Fig. 1A

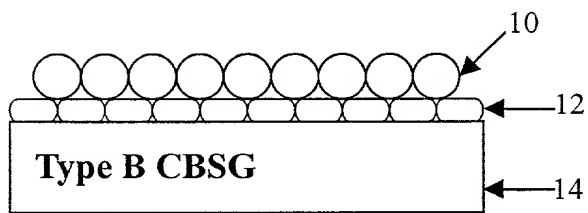


Fig. 1B

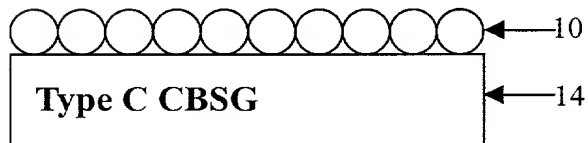


Fig. 1C

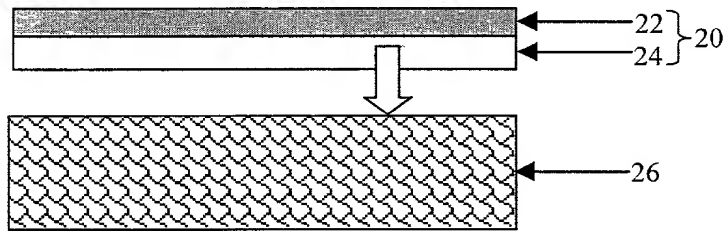


Fig. 2A

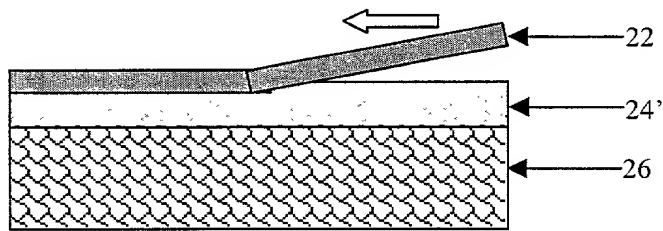


Fig. 2B

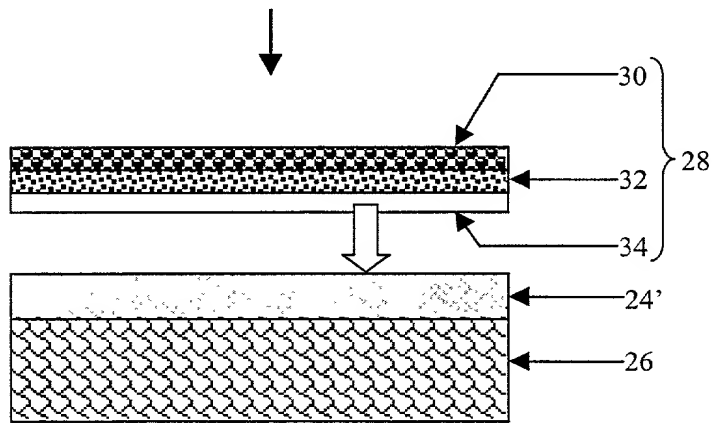


Fig. 2C

**DECLARATION AND POWER OF ATTORNEY**

As a below named inventor, I declare that:

My residence, post office address and citizenship are as stated below next to my name; I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled: **MATERIAL AND METHOD FOR ENGRAFTMENT OF A COMPOSITE BIOCOMPATIBLE SKIN GRAFT ON THE NEODERMIS OF ARTIFICIAL SKIN** the specification of which   X   is attached hereto or        was filed on                      as Application No.                      and was amended on                      (if applicable).

I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56. I claim foreign priority benefits under Title 35, United States Code, Section 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

**POWER OF ATTORNEY:** As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith.

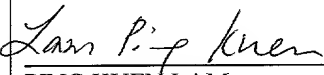
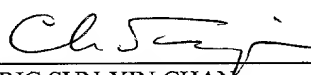
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I further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Signature of Inventor 1  PING KUEN LAM Date 29 July 1999	Signature of Inventor 2  ERIC SUN-YIN CHAN Date July 29, 1999
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